

Disease Note

First report of *Didymella Glomerata* (Corda) and *Penicillium Polonicum* Zaleski on Wheat (*Triticum aestivum* L.) Seeds in Iraq

Ameer Ghani Abd Oun, Mohammed Hamza Abass*

*Department of Plant Protection, College of Agriculture, University of Basrah, Basra, Iraq

Corresponding Author: M H Abass: dr.mha24@yahoo.co.uk

Doi:10.29072/basjs.202301010

ARTICLE INFO

ABSTRACT

Keywords

Didymella glomerata,
morphology,
Penicillium
polonicum,
phylogenetic analysis,
seed- borne.

In the present study, two fungal species, *Didymella glomerata* and *Penicillium polonicum*, were isolated from seeds of four wheat cultivars, named Mahmoodia (MHD), Adena (ADN), Eba 99 (EBA), and Wefia (WAF), and determined by using phenotypic characteristics and molecular sequencing. Molecular diagnosis for both fungi was applied based on internal transcribed spacer primers (ITS1 and ITS4). This is the first record of *D. glomerata* and *P. polonicum* as wheat seed-borne fungi in Iraq.

Received 22 Mar 2023; Received in revised form 12 Apr 2023; Accepted 21 Apr 2023, Published 30 Apr 2023



Introduction

Wheat *Triticum aestivum* L. is one of the most necessary grain crops playing a vital role in attaining global food security [1]. Wheat grains supply more than 18% of calories and 50% of carbohydrates consumed in the world. It also contains a high amount of various phytochemicals, including tannins, alkali, flavonoids, saponins, steroids, terpenoids, and glycosides, that support the conventional system of healthcare for humans [2]. In Iraq, this crop is considered a main staple of nutrition for most if not all population. Moreover, its local production was more than three million tonnes [3]. The wheat crop is attacked by different fungal species including seed-borne fungi whether at pre-harvest or at post-harvest resulting in economic losses in production [4]. Seeds could be passive carriers of fungal pathogens from stores to agrarian areas. Those infected seeds become the source of primary infection when grown in the fields causing seedling damage, seed abortion, seed necrosis, elimination or reduction germination, and seed rot leading to disease development by systemic infections during the growth of crop [5, 6]. Because of the limitation of data on both fungal species, the aim of current report is to determine particularly exact *Didymella glomerata* and *Penicillium polonicum* associated with wheat seeds, as the first record in Iraq. Initially, two species were isolated from sterilized seeds of selected wheat cultivars cultured on full-strength PDA media. Then, phenotypic and molecular characteristics for each fungal genera were identified after purification process of fungal cultures. Identification of the fungal isolates was used to correspond with that mention in related studies of *D. glomerata* [7, 8] and *P. polonicum* [9, 10]. In their research, Valenzuela-Lopez *et al.* [11] focused on the species of the families Didymellaceae and Cucurbitariaceae including *D. glomerata* described phenotypically and molecularly. Moreover, Alidadi *et al.* [6] recorded *D. glomerata* as a phytopathogen and examined its phenotypic and molecular features. Khalil *et al.* [8] confirmed that *P. polonicum* could produce reproductive and vegetative structures isolated from investigated plant samples. They also found that the fungus was mycotoxigenic fungal pathogen being able to synthesize fatal mycotoxin such as citrinin, cyclopiazonic acid and penicillic acid. Chen *et al.* [12] also identified that the fungus could produce Ochratoxin A. Regarding *D. glomerata*, colonies were dark brown at the center and white at edges, fluffy and grew on PDA ranging 74 mm in diameter after nine days at 25°C. The mycelia were brown, soft and loose. Conidia were hyaline, single cell and ellipsoidal measured 3.4-3.7 µm in width and 6.5-6.8 µm in length (Fig.1 A and B). Concerning *P. polonicum*, colonies on PDA were smooth, fluffy and bluish green with white margins reaching



8.2 μm in diameter after nine days at 25°C. Conidiophores were septate and straight with smooth to rough walls. Phialides (conidiogenous cells) were ampulliform. Conidia were sub-globose to globose, smooth walls, and generated in columns. Moreover, conidial measurements were 2.7-3.8 μm in length and 2.4-3.7 μm in width (Fig. 2 A and B).

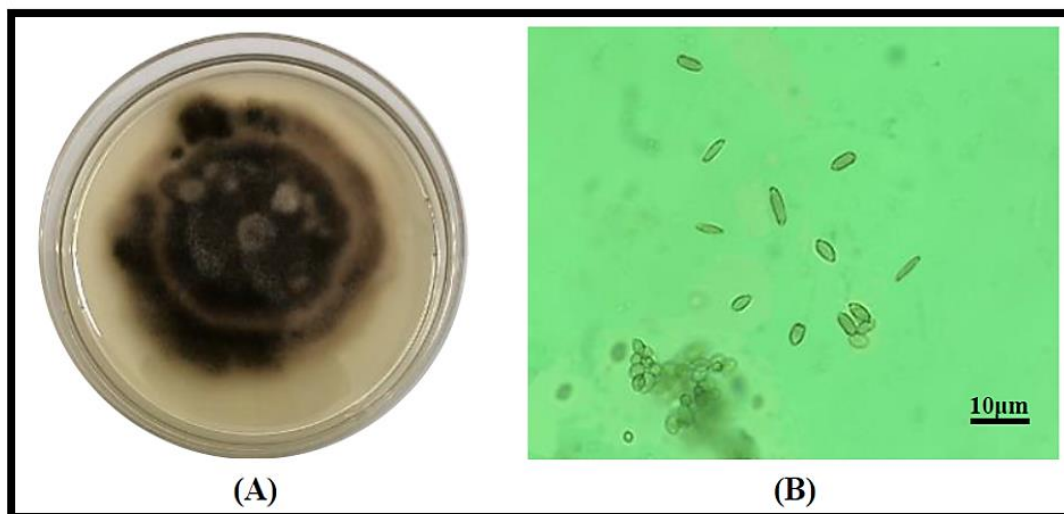


Fig. 1: Morphological traits of *D. glomerata*

(A) Morphology of colony on PDA after nine days, (B) Conidia with a magnification of 40x. Bars: B = 10 μm .

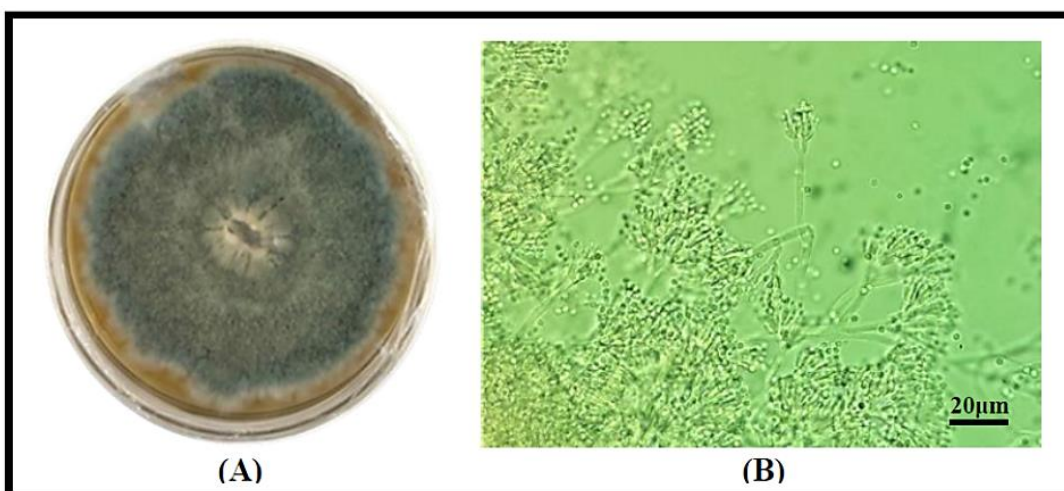


Fig. 2: Morphological traits of *P. polonicum*

(A) Morphology of colony on PDA after nine days, (B) Conidia, conidiophores and phialides with a magnification of 40x. Bars: B = 20 μm .

Both examined fungi were identified molecularly by applying ITS primers (ITS1 and ITS4) [13], the results of phylogenetic analysis revealed the similarity index of 99% for both *D. glomerata* (LT6033041.1) and *P. polonicum* (MT487786.1). The phylogenetic trees were constructed to show the similarities as illustrated in Figure (3 and 4).

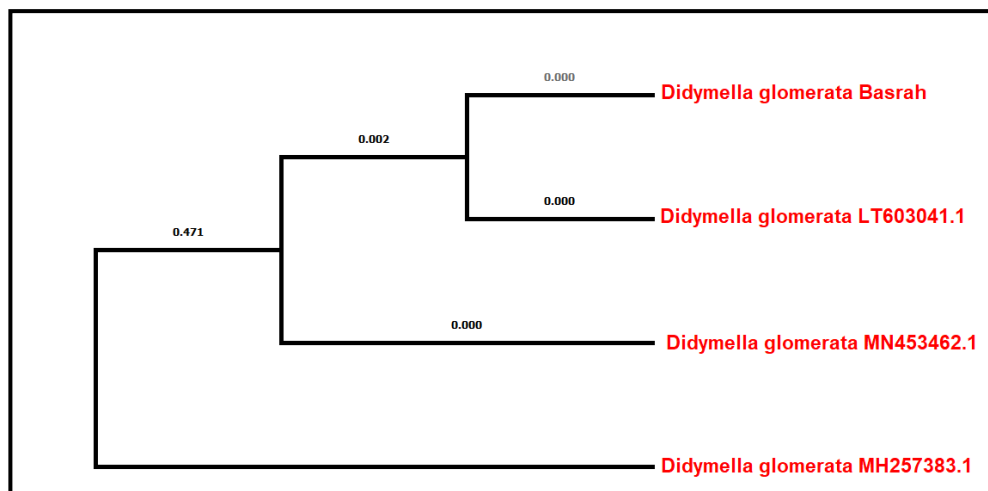


Fig. 3: Phylogenetic analysis of *Didymella glomerata* Basrah isolate.

Maximum likelihood analysis (MLA): phylogenetic tree deduced using Internal Transcribed Spacer (ITS). The nearest three *Didymella glomerata* isolates published in GenBank (<https://www.ncbi.nlm.nih.gov/genbank/samplerecord/>) were used in ClustalW program in MEGA-11 to construct the tree.

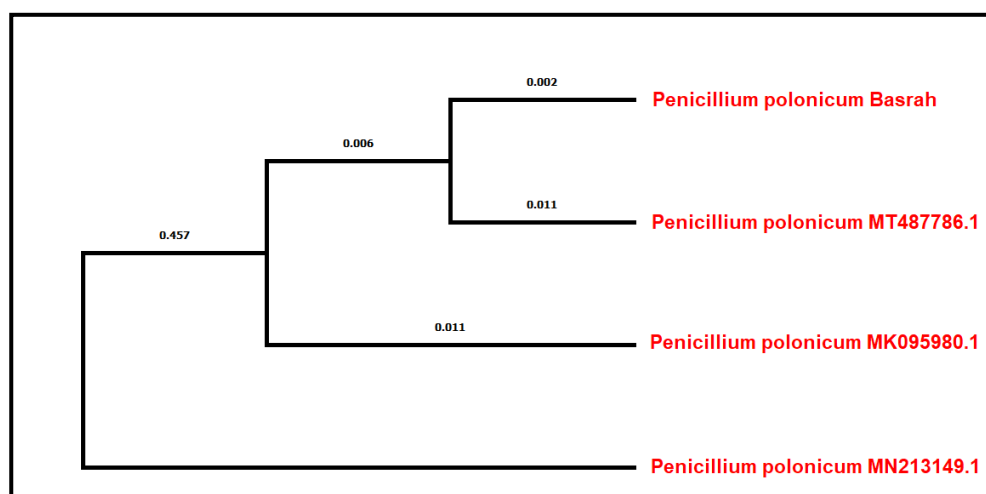


Fig.4: Phylogenetic analysis of *Penicillium polonicum* Basrah isolate.



Maximum likelihood analysis (MLA): phylogenetic tree deduced using Internal Transcribed Spacer (ITS). The nearest three *P. polonicum* isolates published in GenBank (<https://www.ncbi.nlm.nih.gov/genbank/samplerecord/>) were used in ClustalW program in MEGA-11 to construct the tree.

References

- [1] A. M. Khan, M. Khan, H. M. Salman, H. M. Z. U. Ghazali, R. I. Ali, M. Hussain, M. M. Yousaf, Z. Hafeez, M. S. Khawaja, S. A. Alharbi, S. Alfarraj, Detection of seed-borne fungal pathogens associated with wheat (*Triticum aestivum* L.) seeds collected from farmer fields and grain market, J King Saud University-Science, 35(2023)1-7, <https://doi.org/10.1016/j.jksus.2023.102590>
- [2] M. H. Abass, Q. H. Madhi, A. A. Matrood, Identity and prevalence of wheat damping-off fungal pathogens in different fields of Basrah and Maysan provinces, Bulletin of the National Research Centre, 45, (2021) 1-12, <https://doi.org/10.1186/s42269-021-00506-0>
- [3] M. K. Mohammd-Ameen, M. H. Minati, and M. H. Abass, Morphogenetic identification, description and pathogenicity of novel pathogens on Iraqi wheat plant (*Triticum aestivum*) causing head blight and crown rot diseases, Biodiversitas Journal of Biological Diversity, 22(5), (2021), pp. 2999-3005, <https://doi.org/10.13057/biodiv/d220565>
- [4] W. F. Fadhil, A. H. Al-Saadoon, F. M. Al-Moussawi, New records of mycobiota associated with stored wheat and its by-products in Iraq, Biodiversitas J Bio Diversity, 23(2022)3099-3107, <https://doi.org/10.13057/biodiv/d230637>
- [5] S. Sadhasivam, M. Britzi, V. Zakin, M. Kostyukovsky, A. Trostanetsky, E. Quinn, and E. Sionov, Rapid detection and identification of mycotoxigenic fungi and mycotoxins in stored wheat grain, Toxins, 9(2017)1-17, <https://doi.org/10.3390/toxins9100302>
- [6] I. Martin, L. Galvez, L. Guasch, D. Palmero, Fungal pathogens and seed storage in the dry state, Plants, 11(2022)1-25, <https://doi.org/10.3390/plants11223167>
- [7] A. Alidadi, M. Kowsari, M. Javan-Nikkhah, G. S. Jouzani, and M. E. Rastaghi, New pathogenic and endophytic fungal species associated with Persian oak in Iran, Eur J Plant Pathology, 155(2019)1017-1032, <https://doi.org/10.1007/s10658-019-01830-y>
- [8] W. Ma, J. Yang, X. Gao, T. Han, J. Liu, J. Ding, W. Zhao, Y. L. Peng, V. Bhadauria, First Report of *Didymella glomerata* Causing *Didymella* Leaf Blight on Maize. Plant Disease, 106(2022)2522, <https://doi.org/10.1094/PDIS-02-22-0282-PDN>



- [9] A. M. A. Khalil, A. H., Hashem, A. M. Abdelaziz, Occurrence of toxigenic *Penicillium polonicum* in retail green table olives from the Saudi Arabia market, *Biocatalysis Agricul Biotechnolo*, 21 (2019)101314, <https://doi.org/10.1016/j.bcab.2019.101314>
- [10] R. J. Uy, M. Kayamori, and C. Nakashima, Characterization of *Penicillium* Species Isolated from *Dioscorea polystachya* in Hokkaido, Japan, *Mycoscience*, 64(2022)11-18, <https://doi.org/10.47371/mycosci.2022.11.002>
- [11] N. Valenzuela-Lopez, J. F. Cano-Lira, J. Guarro, D. A. Sutton, N. Wiederhold, P. W. Crous, A.M. Stchigel, Coelomycetous Dothideomycetes with emphasis on the families Cucurbitariaceae and Didymellaceae. *Studies in mycology*, 90(2018)1-69, <https://doi.org/10.1016/j.simyco.2017.11.003>
- [12] A. J. Chen, D. Tang, Y. Q. Zhou, B. D. Sun, X. J. Li, L. Z. Wang, W. W. Gao, Identification of ochratoxin A producing fungi associated with fresh and dry liquorice. *PLOS One*, 8(10), (2013), p.78285, <https://doi.org/10.1371/journal.pone.0078285>
- [13] A. N. Ahmed, and M. H. Abass, Disease Note: First Report of *Cladosporium ramotenellum* Schub., Zalar, Crous and Braun, 2007 (Fungi: Dothideomycetes) as a Potential Contaminant of Date Palm Tissue Culture, *Bas J Agricul Sci*, 35(2022)373–375, <https://doi.org/10.37077/25200860.2022.35.2.28>

التسجيل الاول للفطرين *Penicillium polonicum* و *Didymella glomerata* (Corda)

Zaleski على بذور (*Triticum aestivum* L.) الحنطة في العراق

أمير غني عبد عون محمد حمزة عباس*

● قسم وقاية النبات/ كلية الزراعة/ جامعة البصرة/ العراق.

المستخلص

في هذه الدراسة، تم عزل الفطرين *Penicillium polonicum* و *Didymella glomerata* من بذور أربعة أصناف حنطة وهي المحمودية (MHD)، أدنا (ADN)، إباء 99 (EBA) ووفية (WAF)، وتشخيصهما على اساس الصفات المظهرية والمجهريّة وكذلك باستخدام التتابع الجزيئي. وتم تطبيق التشخيص الجزيئي لكلا الفطرين اعتماداً على الواسمات (ITS1 و ITS4) والتي تُعد أول تسجيل لـ *P. polonicum* و *D. glomerata* كفطريات محمولة ببذور الحنطة في العراق.

